# CONJUGATES OF SEVERAL LUPANE, OLEANANE, AND URSANE TRITERPENOIDS WITH THE ANTITUBERCULOSIS DRUG ISONIAZID AND PYRIDINECARBOXALDEHYDES

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Conjugates of betulinic, oleanolic, ursolic, and glycyrrhetic acids and several of their derivatives with the antituberculosis drug isoniazid and 3- and 4-pyridinecarboxaldehydes were synthesized.

**Keywords:** triterpenoids, betulinic, oleanolic, ursolic, glycyrrhetic acids, 28-oxoallobetulone, conjugates, antituberculosis activity, isoniazid.

Betulinic, oleanolic, ursolic, and glycyrrhetic acids have been successfully modified in the last decades to produce new biologically active compounds [1–6] of various structures including hydrazides and hydrazones [4] with antiviral and antitumor activity. It was shown recently that functionalization of the diterpenoid oxo-acid isosteviol by N-containing groups increased its antituberculosis activity, reducing the minimal inhibitory concentrations (MIC) from 50 to  $1.5 \,\mu$ g/mL [7]. Because antituberculosis activity is also typical of triterpenoids, namely betulin [8] and betulinic [8–11], oleanolic [10–13], and ursolic [10, 14] acids, it seemed interesting to perform an analogous investigation of the isoprenoid series by studying the effect of their functionalization by the same *N*-containing groups that were used for isosteviol [7].

Herein we report the synthesis of conjugates of betulinic, oleanolic, ursolic, and glycyrrhetic acids and 28-oxoallobetulone with the antituberculosis drug isoniazid (4-pyridinecarboxylic acid hydrazide) and 3- and 4-pyridinecarboxaldehydes.

Mixed hydrazides 7–9 (Scheme 1) were synthesized via reaction of acid chlorides 4–6, which were prepared from triterpene acids 1–3, with isoniazid by refluxing in CHCl<sub>3</sub> in analogy with the previous work [15]. The target products were isolated by column chromatography in 71–77% yields. The PMR spectra turned out to be sufficiently characteristic to establish their structures. They showed resonances for amide protons at weak field ( $\delta$  6.35 ppm, 8.15, 9.00–9.25, 10.15–10.90). Doubled resonances for the amide protons were noted in 7. This indicated that conformational equilibrium around the C(O)–N bond was occurring in solution. Protons of the pyridine rings of 7–9 appeared as doublets at  $\delta$  7.56–7.68 (H-3', H-5') and 8.50–8.67 (H-2', H-6') ppm. <sup>13</sup>C NMR spectra exhibited resonances for C-28 (C-30) and C-7' at  $\delta$  174.3–176.3 and 161.4–163.7 ppm.



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Pyridinylmethylidenehydrazides 14–19 (Scheme 2) were synthesized via reaction of the hydrazides of triterpene acids 10–13 with 3- or 4-pyridinecarboxyaldehydes by refluxing in EtOH in analogy with the previous work [16]. The products were isolated by column chromatography in 51–59% yields. Resonances of protons of C(28 or 30)ONH- and =C(7')H-groups were observed at  $\delta$  8.20–10.40 and 8.00–8.70 ppm, respectively, in PMR spectra of 14–19. The pyridine protons of 14, 15, 17, and 18 were observed at weak field at 7.31–7.35 ppm (H-5', multiplet), 8.14–8.22 (H-4', doublet), 8.45–8.55 (H-6', doublet), and 8.65–9.35 (H-2', singlet). Protons H-3', H-5' and H-2', H-6' resonated in spectra of 16 and 19 as doublets at  $\delta$  7.39–7.50 and 8.63–8.70 ppm, respectively. <sup>13</sup>C NMR spectra of 18–27 contained resonances of C-28 (C-30) and C-7' at  $\delta$  172.9–173.4 and 142.7–150.6 ppm, respectively.



i. 3- or 4-pyridinecarboxaldehyde, EtOH, reflux

#### Scheme 2

Compounds 20–23, 25, 26, and 28–31 (Scheme 3) with a pyridinylmethylidene substituent in the C-2 position were prepared via reaction of 3-oxotriterpenoids 1–3, 24, and 27 with 3- or 4-pyridinecarboxaldehydes in basic EtOH in analogy with the previous work [17] in 88–95% yields (after recrystallization). PMR spectra of 20–23, 25, 26, and 28–31 had the resonance for the methylidene proton at  $\delta$  6.85–8.50 ppm. The pyridine protons in spectra of 20, 22, 25, 28, and 30 resonated as multiplets at  $\delta$  7.30–7.41 ppm (H-5'), doublets at 7.70–8.55 (H-4'), doublets at 7.72–8.62 (H-6'), and singlets at 8.65–8.70 ppm (H-2'). Spectra of 21, 23, 26, 29, and 31 showed doublets at  $\delta$  7.06–7.41 and 8.45–8.65 ppm corresponding to protons H-3', H-5' and H-2', H-6'. <sup>13</sup>C NMR spectra of 20–23, 25, 26, and 28–31 showed resonances of C-7' at  $\delta$  136.2–144.1 ppm; of the pyridine C atoms, at weak field in the range from  $\delta$  121.8 to 181.9 ppm.

The antituberculosis activity of the synthesized compounds was evaluated preliminarily at the National Institute of Allergy and Infectious Diseases (USA, www.niaid-aacf.org). It was found that conjugates 7–9, 15–18, 20, 22, 23, 25, 26, 30, and 31 did not inhibit growth of *Mycobacterium tuberculosis* (strain  $H_{37}Rv$  *in vitro*) at MIC values <10 µg/mL. Unfortunately, the conditions of the preliminary screening did not provide for a study of the antituberculosis activity at MIC values >10 µg/mL. We were unable in this stage to evaluate the effect of covalent binding of isoniazid and pyridinecarboxaldehydes with triterpenoid betulinic, oleanolic, ursolic, and glycyrrhetic acids on the known inhibiting capability of the last (MIC = 12.5–100 µg/mL [8–14]). Therefore, syntheses of conjugates of triterpenoid metabolites with antituberculosis activity with synthetic mycostatics and studies of their antituberculosis activity will be continued.

Thus, we synthesized conjugates of several triterpenoid acids and their derivatives with the antituberculosis drug isoniazid and pyridinecarboxaldehydes in addition to 2-pyridinylmethylidene derivatives of 28-oxoallobetulone.





#### **EXPERIMENTAL**

PMR and <sup>13</sup>C NMR spectra were recorded in  $\text{CDCl}_3$  with TMS internal standard on a Bruker AM-300 spectrometer (300 and 75.5 MHz, respectively). Melting points were determined on a Boetius microstage. Optical density was measured in a 1-dm tube on a Perkin–Elmer 241 MC polarimeter. TLC analysis used Sorbil plates (ZAO Sorbpolimer, Russia) and solvent system  $\text{CHCl}_3$ :EtOAc (40:1). Compounds were detected by  $\text{H}_2\text{SO}_4$  solution (10%) with subsequent heating at 100–120°C for 2–3 min. We used oleanolic and ursolic acids and 3- and 4-pyridinecarboxaldehydes (Aldrich Chemical Co.). Betulonic acid (1) was synthesized from betulin by the literature method [18]. Ursolic and oleanolic acids were oxidized by Jones reagent in acetone to the 3-oxo derivatives 2 and 24 in analogy with previous work [18]. 3-Oxoglycyrrhetic acid (3) and 28-oxoallobetulone (27) were prepared as before [19, 20]. Constants of the starting compounds agreed with those published. Acid chlorides 4–6 were prepared by the standard method [15] via reaction with oxalylchloride in CHCl<sub>3</sub>. Triterpene acid hydrazides 10–13 were synthesized via reaction of their acid chlorides with hydrazine hydrate in CHCl<sub>3</sub> as before [16]. Hydrazides of 3- and 4-pyridinecarboxylic acids were prepared by the literature method [21].

**Synthesis of 7–9.** A solution of freshly prepared acid chloride **4–6** (1 mmol) in anhydrous  $CHCl_3$  (20 mL) was treated with the hydrazide of 4-pyridinecarboxylic acid (1.5 mmol), refluxed until the reaction was complete (TLC monitoring), and diluted with HCl solution (30 mL, 5%). The precipitate was filtered off, washed with H<sub>2</sub>O, and dried. The product was chromatographed over a column with elution successively by  $C_6H_6$  and  $CHCl_3$ .

17β-(4-Pyridinoylhydrazinocarbonyl)-3-oxolup-20(29)-ene (7). Yield 0.42 g (73%), mp 192–194°C,  $[\alpha]_D^{20}$  +8° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>51</sub>O<sub>3</sub>N<sub>3</sub> (MW 573.81).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.80, 0.89, 0.90, 0.95, 0.95, 1.59 (18H, 6s, 6CH<sub>3</sub>), 1.10–2.50 (24H, m, CH<sub>2</sub>, CH), 2.90 (1H, m, H-19), 4.50 (1H, s, H-29a), 4.60 (1H, s, H-29b), 7.60 (2H, d, J = 5.3, H-3', H-5'), 8.50 (2H, d, J = 2.4, H-2', H-6'), 8.15 (2H, br. signal, C<sup>28</sup>ON<u>H</u>, C<sup>7'</sup>ON<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 14.5, 15.9, 19.3, 19.6, 21.0, 21.4, 25.6, 26.6, 29.4, 30.8, 33.1, 33.5, 34.1, 36.9, 37.7, 38.4, 39.0, 39.6, 40.7, 42.5, 46.7, 47.3, 49.9, 50.3, 55.0, 55.4, 109.7 (C-29), 121.2 (C-3'), 121.2 (C-5'), 138.9 (C-4'), 149.0 (C-20), 150.3 (C-2'), 150.3 (C-6'), 163.6 (C-7'), 176.3 ( $\underline{C}^{28}$ ONH), 218.1 (C-3).

17β-(4-Pyridinoylhydrazinocarbonyl)-3-oxours-12-ene (8). Yield 0.44 g (77%), mp 168–170°C,  $[\alpha]_D^{20}$ +27° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>51</sub>O<sub>3</sub>N<sub>3</sub> (MW 573.81).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.77, 0.86, 0.88, 0.93, 1.01, 1.05, 1.09 (21H, 7s, 7CH<sub>3</sub>), 1.15–2.58 (23H, m, CH<sub>2</sub>, CH), 5.50 (1H, s, H-12), 7.68 (2H, d, J = 3.3, H-3', H-5'), 8.67 (2H, d, J = 3.3, H-2', H-6'), 9.25 (1H, br. signal, C<sup>28</sup>ON<u>H</u>), 10.90 (1H, br. signal, C<sup>7</sup>ON<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 15.3, 16.4, 17.1, 19.6, 21.1, 21.5, 23.3, 23.6, 24.9, 26.5, 27.8, 30.7, 32.2, 34.1, 36.6, 36.9, 38.9, 39.3, 39.5, 39.5, 42.4, 46.8, 47.4, 47.7, 52.9, 55.2, 121.2 (C-12), 127.1 (C-3'), 127.1 (C-5'), 138.6 (C-4'), 138.9 (C-13), 150.3 (C-2'), 150.3 (C-6'), 161.4 (C-7'), 174.3 (C-28), 217.4 (C-3).

**20** $\beta$ -(4-Pyridinoylhydrazinocarbonyl)-3,11-dioxoolean-12-ene (9). Yield 0.41 g (71%), mp 197°C,  $[\alpha]_D^{20}$  +129° (*c* 0.1, CHCl<sub>3</sub>),  $C_{36}H_{49}O_4N_3$  (MW 587.79).

PMR spectrum (CDCl<sub>3</sub>,  $\delta$ , ppm, J/Hz): 0.62, 0.95, 0.99, 1.01, 1.13, 1.15, 1.25 (21H, 7s, 7CH<sub>3</sub>), 1.20–2.60 (20H, m, CH<sub>2</sub>, CH), 2.80 (1H, m, H-18), 5.65 (1H, s, H-12), 7.56 (2H, d, J = 5.7, H-3', H-5'), 8.57 (2H, d, J = 5.1, H-2', H-6'), 9.00 (1H, br. signal, C<sup>7</sup>ON<u>H</u>), 10.15 (1H, br. signal, C<sup>30</sup>ON<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 15.6, 18.4, 18.7, 19.5, 21.3, 23.2, 26.3, 26.5, 28.4, 28.9, 31.2, 31.7, 32.0, 34.1, 36.6, 37.1, 39.7, 41.5, 43.2, 43.3, 45.2, 47.7, 47.9, 55.3, 61.0, 121.1 (C-3'), 121.1 (C-5'), 128.1 (C-12), 138.6 (C-4'), 150.4 (C-2'), 150.4 (C-6'), 163.7 (C-7'), 170.2 (C-13), 175.4 (<u>C</u><sup>30</sup>ONH), 199.8 (C-11), 217.1 (C-3).

Synthesis of 14–19. A solution of the acid hydrazides 10-13 (1 mmol) in EtOH (30 mL) was treated with 3- or 4-pyridinecarboxaldehyde (1.3 mmol), refluxed until the reaction was complete (TLC monitoring), and poured into HCl solution (100 mL, 5%). The precipitate was filtered off, washed with H<sub>2</sub>O until neutral, and dried. The product was chromatographed over a column with elution successively by CHCl<sub>3</sub> and CHCl<sub>3</sub>:MeOH (50:1).

17β-(Pyridin-3-ylmethylidenehydrazinocarbonyl)-3β-hydroxylup-20(29)-ene (14). Yield 0.28 g (51%), mp 143–145°C,  $[\alpha]_D^{20}$  –1° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>53</sub>O<sub>2</sub>N<sub>3</sub> (MW 559.82).

PMR spectrum (CDCl<sub>3</sub>,  $\delta$ , ppm, J/Hz): 0.69, 0.78, 0.90, 0.92, 0.96 (15H, 5s, 5CH<sub>3</sub>), 1.10–2.30 (25H, m, CH<sub>2</sub>, CH), 1.65 (3H, s, C<sup>30</sup><u>H</u><sub>3</sub>), 2.70 (1H, m, H-3), 4.30 (1H, br. signal,  $-O\underline{H}$ ), 4.56 (1H, s, H-29a), 4.69 (1H, s, H-29b), 7.35 (1H, dd, J = 7.85, 7.90, H-5'), 8.15 (1H, s, H-7'), 8.22 (1H, d, J = 7.90, H-4'), 8.45 (1H, d, J = 7.85, H-6'), 8.65 (1H, s, H-2'), 10.10 (1H, br. signal, C<sup>28</sup>ON<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 14.3, 15.0, 15.9, 17.5, 18.0, 19.0, 20.8, 25.4, 26.7, 27.6, 29.3, 30.4, 32.2, 33.7, 34.2, 36.9, 37.1, 38.6, 40.5, 42.1, 46.1, 50.3, 50.5, 54.9, 55.3, 57.2, 78.4 (C-3), 109.1 (C-29), 123.9 (C-5'), 130.9 (C-3'), 134.0 (C-4'), 142.7 (C-7'), 148.4 (C-20), 149.6 (C-2'), 150.5 (C-6'), 174.0 (<u>C</u>ONH).

17β-(Pyridin-3-ylmethylidenehydrazinocarbonyl)-3-hydroxyiminolup-20(29)-ene (15). Yield 0.31 g (55%), mp 195–197°C,  $[α]_D^{20}$ +5° (c 0.1, CHCl<sub>3</sub>),  $C_{36}H_{52}O_2N_4$  (MW 572.82).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm): 0.85, 0.95, 1.10, 1.35, 1.45, 1.70 (18H, 6s, 6CH<sub>3</sub>), 1.80–3.20 (25H, m, CH<sub>2</sub>, CH), 4.60 (1H, s, H-29a), 4.70 (1H, s, H-29b), 7.30-9.00 (5H, m, H-2', H-4', H-5', H-6', H-7'), 9.92 (2H, br. signal, CON<u>H</u>, NO<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 11.6, 14.6, 17.0, 19.4, 21.1, 23.0, 26.6, 27.1, 29.7, 29.8, 30.8, 34.2, 36.9, 37.2, 38.8, 40.3, 40.8, 42.5, 42.6, 43.7, 46.3, 47.4, 50.1, 50.4, 55.5, 55.9, 109.6 (C-29), 123.9 (C-5'), 130.5 (C-3'), 134.4 (C-4'), 143.6 (C-7'), 146.1 (C-20), 150.6 (C-6'), 159.8 (C-2'), 166.9 (C-3), 173.4 (<u>C</u>ONH).

17β-(Pyridin-4-ylmethylidenehydrazinocarbonyl)-3-hydroxyiminolup-20(29)-ene (16). Yield 0.33 g (58%), mp 185–190°C,  $[\alpha]_D^{20}$  +8° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>52</sub>O<sub>2</sub>N<sub>4</sub> (MW 572.82).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm): 0.92, 0.98, 1.08, 1.31, 1.41, 1.62 (18H, 6s, 6CH<sub>3</sub>), 1.20–3.15 (25H, m, CH<sub>2</sub>, CH), 4.60 (1H, s, H-29a), 4.70 (1H, s, H-29b), 7.50-8.71 (5H, m, H-2', H-3', H-5', H-6', H-7'), 10.00 (2H, br. signal, CON<u>H</u>, NO<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 14.6, 15.9, 16.2, 17.4, 19.6, 21.0, 22.0, 22.9, 25.6, 26.6, 29.6, 29.8, 30.4, 30.7, 33.7, 34.1, 36.9, 39.6, 40.7, 42.3, 44.2, 46.2, 47.3, 50.1, 55.0, 58.0, 109.4 (C-29), 121.1 (C-3'), 121.1 (C-5'), 143.6 (C-7'), 144.3 (C-4'), 150.1 (C-2'), 150.1 (C-6'), 150.5 (C-20), 166.9 (C-3), 173.4 (<u>C</u>ONH).

17β-(Pyridin-3-ylmethylidenehydrazinocarbonyl)-3β-hydroxyurs-12-ene (17). Yield 0.33 g (59%), mp 186–188°C,  $[\alpha]_D^{20}$  -2° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>53</sub>O<sub>2</sub>N<sub>3</sub> (MW 559.82).

 $\begin{array}{l} PMR \ spectrum \ (CDCl_3, \delta, ppm, J/Hz): \ 0.70, \ 0.72, \ 0.84, \ 0.86, \ 0.88, \ 0.95, \ 1.10 \ (21H, \ 7s, \ 7CH_3), \ 1.20-2.10 \ (22H, \ m, \ CH_2, \ CH), \ 2.70 \ (1H, \ d, \ J = 11.5, \ H-18), \ 3.20 \ (1H, \ dd, \ J = 11.0, \ 4.1, \ H-3), \ 5.48 \ (1H, \ s, \ H-12), \ 7.31 \ (2H, \ m, \ H-5', \ OH), \ 8.14 \ (1H, \ d, \ J = 7.8, \ H-4'), \ 8.20 \ (1H, \ s, \ CON\underline{H}), \ 8.55 \ (1H, \ d, \ J = 4.1, \ H-6'), \ 8.70 \ (1H, \ s, \ H-7'), \ 9.35 \ (1H, \ s, \ H-2'). \end{array}$ 

<sup>13</sup>C NMR spectrum (δ, ppm): 15.5, 15.6, 16.9, 17.2, 18.2, 21.2 23.4, 23.6, 25.1, 27.2, 27.9, 28.2, 30.8, 32.6, 36.9, 36.9, 38.7, 38.8, 39.0, 39.6, 39.7, 42.5, 47.5, 48.1, 53.4, 55.1, 78.8 (C-3), 123.7 (C-5'), 128.3 (C-12), 130.2 (C-3'), 133.8 (C-4'), 140.2 (C-13), 144.3 (C-7'), 149.4 (C-2'), 150.9 (C-6'), 174.9 (<u>C</u>ONH).

**20**β-(Pyridin-3-ylmethylidenehydrazinocarbonyl)-11-oxo-3β-hydroxyolean-12-ene (18). Yield 0.32 g (56%), mp 166–168°C,  $[\alpha]_D^{20}$ +103° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>51</sub>O<sub>3</sub>N<sub>3</sub> (MW 573.81).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.80, 0.80, 1.00, 1.10, 1.10, 1.20, 1.40 (21H, 7s, 7CH<sub>3</sub>), 1.20–2.40 (20H, m, CH<sub>2</sub>, CH), 2.70 (1H, d, J = 11.5, H-18), 3.20 (1H, br. signal, H-3), 5.78 (1H, s, H-12), 8.00–8.60 (5H, m, H-2', H-4', H-5', H-6', H-7'), 10.4 (2H, br. signal,  $-O\underline{H}$ , CON<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 15.5, 16.2, 17.4, 18.3, 18.6, 23.3, 26.3, 27.1, 28.1, 28.3, 29.0, 31.2, 31.8, 32.7, 37.0, 37.4, 39.1, 39.2, 41.1, 43.2, 45.4, 48.0, 54.9, 58.0, 61.8, 78.6 (C-3), 123.7 (C-5'), 128.3 (C-12), 130.1 (C-3'), 133.7 (C-4'), 144.5 (C-7'), 149.0 (C-2'), 150.5 (C-6'), 169.6 (C-13), 172.9 (<u>C</u>ONH), 200.3 (C-11).

**20**β-(**Pyridin-4-ylmethylidenehydrazinocarbonyl)-11-oxo-3**β-hydroxyolean-12-ene (19). Yield 0.34 g (59%), mp 201°C,  $[\alpha]_D^{20}$  +162° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>51</sub>O<sub>3</sub>N<sub>3</sub> (MW 573.81).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm): 0.78, 1.00, 1.05, 1.10, 1.21, 1.29, 1.34 (21H, 7s, 7CH<sub>3</sub>), 1.40–2.85 (21H, m, CH<sub>2</sub>, CH), 5.78 (1H, s, H-12), 7.51 (2H, br.s, H-3', H-5'), 8.21 (1H, s, H-7'), 8.52 (2H, br. signal, H-2', H-6'), 10.26 (1H, br. signal, CON<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 15.5, 18.4, 18.6, 21.3, 23.2, 26.3, 28.4, 28.5, 28.9, 31.2, 31.7, 32.0, 34.1, 36.6, 37.3, 39.7, 40.9, 43.1, 43.3, 43.5, 45.1, 47.7, 47.9, 55.3, 61.0, 121.4 (C-3'), 121.4 (C-5'), 128.2 (C-12), 141.6 (C-4'), 145.0 (C-7'), 149.9 (C-2'), 149.9 (C-6'), 170.3 (C-13), 173.0 (<u>C</u>ONH), 199.9 (C-11), 217.2 (C-3).

Synthesis of 20–23, 25, 26, 28–31. A solution of 1–3, 24, or 27 (1 mmol) in EtOH (20 mL) was stirred, treated with 3- or 4-pyridinecarboxaldheyde (1.5 mmol) and KOH solution (2.5 mL, 40%) in EtOH, left for 1 d, and poured into HCl solution (100 mL, 5%). The precipitate was filtered off, washed with H<sub>2</sub>O until neutral, dried, and crystallized from EtOH (MeOH).

**3-Oxo-2-(pyridin-3-ylmethylideno)-lup-20(29)-en-28-oic Acid (20).** Yield 0.48 g (89%), mp 183°C,  $[\alpha]_D^{20}$  +17° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>49</sub>O<sub>3</sub>N (MW 543.78).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.97, 1.02, 1.12, 1.14, 1.46, 1.72 (18H, 6s, 6CH<sub>3</sub>), 1.20–2.40 (21H, m, CH<sub>2</sub>, CH), 3.00 (2H, m, H-19, H-1a), 4.61 (1H, s, H-29a), 4.71 (1H, s, H-29b), 7.41 (2H, m, H-5', H-7'), 7.72 (1H, d, J = 7.88, H-4'), 8.54 (1H, d, J = 4.8, H-6'), 8.69 (1H, s, H-2'), 10.80 (1H, br. signal, COO<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm); 14.5, 15.5, 15.8, 19.4, 20.3, 21.6, 22.3, 25.5, 29.4, 29.6, 30.5, 32.1, 33.0, 36.5, 37.0, 38.4, 40.5, 42.5, 44.4, 45.2, 46.8, 48.4, 49.1, 52.8, 56.3, 109.7 (C-29), 123.5 (C-5'), 132.0 (C-3'), 133.1 (C-4'), 136.6 (C-7'), 137.2 (C-2), 148.4 (C-2'), 150.5 (C-6'), 150.6 (C-20), 181.3 (<u>C</u>OOH), 207.7 (C-3).

**3-Oxo-2-(pyridin-4-ylmethylideno)-lup-20(29)-en-28-oic Acid (21).** Yield 0.49 g (90%), mp 183°C,  $[\alpha]_D^{20}$  +29° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>49</sub>O<sub>3</sub>N (MW 543.78).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.96, 0.99, 1.08, 1.11, 1.43, 1.69 (18H, 6s, 6CH<sub>3</sub>), 1.10–2.40 (20H, m, CH<sub>2</sub>, CH), 2.20 (1H, d,  $J_{gem} = 16.9$ , H-1b), 3.00 (2H, dd,  $J_{gem} = 16.9$ , 8.7, H-1a, H-19), 4.62 (1H, s, H-29a), 4.72 (1H, s, H-29b), 7.25 (1H, s, H-7'), 7.30 (2H, d, J = 5.3, H-3', H-5'), 8.63 (2H, d, J = 5.3, H-2', H-6'), 10.80 (1H, br. signal, COO<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 14.7, 15.6, 15.9, 19.6, 20.4, 21.8, 22.4, 25.7, 29.3, 29.8, 30.8, 32.3, 33.1, 36.7, 37.2, 38.5, 40.6, 42.6, 44.4, 45.5, 47.0, 48.5, 49.3, 53.0, 56.4, 109.7 (C-29), 124.4 (C-3'), 124.4 (C-5'), 133.6 (C-2), 139.0 (C-4'), 144.4 (C-7'), 149.1 (C-2'), 149.1 (C-6'), 150.7 (C-20), 180.8 (<u>C</u>OOH), 207.9 (C-3).

**3-Oxo-2-(pyridin-3-ylmethylideno)-urs-12-en-28-oic Acid (22).** Yield 0.49 g (91%), mp 139–145°C,  $[\alpha]_D^{20}$  –18° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>49</sub>O<sub>3</sub>N (MW 543.78).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.81, 0.85, 0.89, 0.91, 0.96, 1.13, 1.13 (21H, 7s, 7CH<sub>3</sub>), 1.15–2.35 (20H, m, CH<sub>2</sub>, CH), 2.95 (1H, d,  $J_{gem}$  = 16.3, H-1a), 5.25 (1H, s, H-12), 7.35 (1H, dd, J = 4.8, 8.0, H-5'), 7.45 (1H, s, H-7'), 7.72 (1H, d, J = 8.0, H-6'), 8.55 (1H, d, J = 3.5, H-4'), 8.70 (1H, s, H-2'), 11.0 (1H, br. signal, COO<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 15.5, 16.7, 17.1, 20.3, 21.2, 22.7, 23.4, 23.5, 24.1, 28.0, 29.6, 30.7, 32.1, 36.3, 36.7, 38.9, 39.2, 39.4, 42.2, 44.1, 45.2, 45.3, 48.0, 52.8, 53.1, 123.6 (C-5'), 125.2 (C-12), 132.0 (C-3'), 133.4 (C-4'), 136.2 (C-7'), 137.5 (C-2), 138.3 (C-13), 148.5 (C-2'), 150.4 (C-6'), 183.0 (<u>C</u>OOH), 207.3 (C-3).

**3-Oxo-2-(pyridin-4-ylmethylideno)-urs-12-en-28-oic Acid (23).** Yield 0.50 g (92%), mp 207°C,  $[\alpha]_D^{20}$  –5° (*c* 0.1, CHCl<sub>2</sub>), C<sub>36</sub>H<sub>49</sub>O<sub>3</sub>N (MW 543.78).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.76, 0.79, 0.83, 0.85, 0.90, 1.60, 1.60 (21H, 7s, 7CH<sub>3</sub>), 1.10–2.30 (20H, m, CH<sub>2</sub>, CH), 2.90 (1H, d,  $J_{gem}$  = 16.5, H-1a), 5.20 (1H, s, H-12), 7.20 (2H, d, J = 4.8, H-3', H-5'), 7.40 (1H, s, H-7'), 8.60 (2H, d, J = 4.8, H-2', H-6'), 11.0 (1H, br. signal, COO<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 15.5, 16.7, 17.1, 20.2, 21.2, 22.7, 23.5, 23.5, 24.2, 28.0, 29.4, 30.7, 32.1, 36.4, 36.7, 38.8, 39.2, 39.4, 42.2, 43.9, 45.2, 45.4, 48.0, 52.8, 53.2, 124.2 (C-3'), 124.2 (C-5'), 125.1 (C-12), 133.9 (C-4'), 138.2 (C-13), 138.2 (C-2), 144.1 (C-7'), 149.3 (C-2'), 149.3 (C-6'), 182.7 (<u>C</u>OOH), 207.3 (C-3).

**3-Oxo-2-(pyridin-3-ylmethylideno)-olean-12-en-28-oic Acid (25).** Yield 0.52 g (95%), mp 192–194°C,  $[\alpha]_D^{20}$ +44° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>49</sub>O<sub>3</sub>N (MW 543.78).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.89, 0.91, 0.95, 1.05, 1.10, 1.12, 1.38 (21H, 7s, 7CH<sub>3</sub>), 1.20–2.50 (18H, m, CH<sub>2</sub>, CH), 2.24 (1H, d,  $J_{gem} = 16.8$ , H-1b), 3.00 (1H, d,  $J_{gem} = 16.8$ , H-1a), 4.20 (1H, m, H-18), 5.69 (1H, s, H-12), 7.35 (1H, d, J = 7.7, H-5'), 7.66 (1H, s, H-7'), 7.89 (1H, d, J = 7.7, H-4'), 8.50 (1H, s, H-2'), 8.62 (1H, s, H-6'), 11.0 (1H, br. signal, COO<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 15.3, 18.4, 19.4, 22.5, 22.8, 23.4, 23.4, 27.9, 29.5, 30.7, 31.6, 32.8, 33.7, 36.4, 41.6, 41.9, 43.0, 43.7, 44.6, 44.8, 45.5, 45.9, 46.0, 53.4, 59.3, 123.7 (C-12), 127.8 (C-5'), 132.5 (C-3'), 137.0 (C-4'), 137.2 (C-7'), 147.8 (C-2), 150.7 (C-13), 170.0 (C-2'), 181.9 (C-6'), 199.5 (<u>C</u>OOH), 207.1 (C-3).

**3-Oxo-2-(pyridin-4-ylmethylideno)-olean-12-en-28-oic Acid (26).** Yield 0.48 g (88%), mp 191–193°C,  $[\alpha]_D^{20}$  +5° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>49</sub>O<sub>3</sub>N (MW 543.78).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.80, 0.83, 0.90, 0.92, 1.12, 1.14, 1.18 (21H, 7s, 7CH<sub>3</sub>), 1.00–2.10 (18H, m, CH<sub>2</sub>, CH), 2.27 (1H, d,  $J_{gem} = 16.8$ , H-1b), 2.88 (1H, m, H-18), 2.94 (1H, d,  $J_{gem} = 16.8$ , H-1a), 5.30 (1H, s, H-12), 7.25 (2H, d, J = 5.8, H-3', H-5'), 7.35 (1H, s, H-7'), 8.61 (2H, d, J = 5.8, H-2', H-6'), 11.0 (1H, br. signal, COO<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 15.2, 16.5, 20.2, 20.5, 22.6, 23.0, 23.5, 25.6, 27.7, 29.5, 30.7, 31.8, 32.4, 33.0, 33.8, 36.3, 39.1, 41.2, 41.9, 44.0, 45.3, 45.9, 46.5, 49.2, 53.0, 121.8 (C-3'), 121.8 (C-52), 124.3 (C-12), 130.6 (C-2), 133.7 (C-4'), 138.3 (C-13), 144.1 (C-7'), 149.1 (C-2'), 149.1 (C-6'), 182.8 (<u>C</u>OOH), 207.3 (C-3).

**3,28-Dioxo-2-(pyridin-3-ylmethylideno)-19** $\beta$ **,28-epoxyoleanane (28).** Yield 0.49 g (90%), mp 154–156°C,  $[\alpha]_D^{20}$  +22° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>49</sub>O<sub>3</sub>N (MW 541.76).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.78, 0.88, 0.91, 0.93, 0.99, 1.99, 1.13 (21H, 7s, 7CH<sub>3</sub>), 1.10–2.15 (20H, m, CH<sub>2</sub>, CH), 2.21 (1H, d,  $J_{gem} = 16.1$ , H-1b), 3.00 (1H, d,  $J_{gem} = 16.2$ , H-1a), 3.91 (1H, m, H-19), 7.30 (1H, m, H-5'), 7.40 (1H, s, H-7'), 7.70 (1H, d, J = 8.0, H-4'), 8.51 (1H, d, J = 4.1, H-6'), 8.66 (1H, s, H-2').

<sup>13</sup>C NMR spectrum (δ, ppm): 13.6, 15.1, 16.2, 20.3, 21.7, 22.4, 24.0, 25.6, 26.6, 27.9, 28.8, 29.5, 32.0, 32.4, 32.5, 33.6, 36.2, 36.7, 40.1, 40.4, 44.8, 45.3, 46.1, 46.7, 49.1, 53.0, 86.0 (C-19), 123.4 (C-5'), 131.9 (C-3'), 133.6 (C-4'), 136.4 (C-7'), 137.1 (C-2), 149.1 (C-2'), 150.9 (C-6'), 179.7 (C-28), 207.6 (C-3).

**3,28-Dioxo-2-(pyridin-4-ylmethylideno)-19** $\beta$ **,28-epoxyoleanane (29).** Yield 0.49 g (92%), mp 247–250°C,  $[\alpha]_D^{20}$  +109° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>49</sub>O<sub>3</sub>N (MW 541.76).

PMR spectrum (CDCl<sub>3</sub>,  $\delta$ , ppm, J/Hz): 0.88, 0.94, 0.96, 0.97, 1.01, 1.03, 1.05 (21H, 7s, 7CH<sub>3</sub>), 1.10–1.90 (20H, m, CH<sub>2</sub>, CH), 3.35 (1H, d, J = 14.9, H-1a), 3.58 (1H, d, J = 14.7, H-1b), 3.92 (1H, s, H-19), 6.85 (1H, s, H-7'), 7.06 (2H, d, J = 5.8, H-3', H-5'), 8.45 (2H, d, J = 5.8, H-2', H-6').

<sup>13</sup>C NMR spectrum (δ, ppm): 13.6, 16.3, 19.1, 20.1, 21.4, 21.5, 24.0, 25.6, 26.4, 27.8, 28.3, 28.8, 32.0, 32.4, 33.1, 33.6, 36.2, 36.3, 39.2, 41.5, 44.7, 45.5, 46.1, 46.7, 49.3, 53.4, 85.9 (C-19), 124.1 (C-3'), 124.1 (C-5'), 133.7 (C-4'), 149.3 (C-7'), 149.7 (C-2'), 149.7 (C-6'), 156.4 (C-2), 179.6 (C-28), 204.4 (C-3).

**3,11-Dioxo-2-(pyridine-3-ylmethylideno)-olean-12-en-30-oic Acid (30).** Yield 0.51 g (91%), mp 140–143°C,  $[\alpha]_D^{20}$  +94° (*c* 0.1, CHCl<sub>3</sub>),  $C_{36}H_{47}O_4N$  (MW 557.76).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.81, 1.10, 1.15, 1.18, 1.21, 1.39, 1.50 (21H, 7s, 7CH<sub>3</sub>), 1.35–2.50 (18H, m, CH<sub>2</sub>, CH), 4.20 (1H, d,  $J_{gem}$  = 16.5, H-1a), 5.79 (1H, s, H-12), 7.39 (1H, dd, J = 4.8, 8.0, H-5'), 7.42 (1H, s, H-7'), 7.92 (1H, d, J = 8.0, H-6'), 8.54 (1H, d, J = 3.5, H-4'), 8.70 (1H, s, H-2'), 11.0 (1H, br. signal, COO<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 15.3, 17.9, 19.4, 22.4, 23.1, 26.3, 26.4, 28.5, 29.4, 29.5, 30.9, 31.3, 31.8, 36.2, 37.6, 41.1, 43.3, 43.6, 44.2, 44.9, 45.4, 48.2, 53.3, 59.1, 123.9 (C-5'), 128.3 (C-12), 131.9 (C-3'), 132.5 (C-4'), 137.3 (C-7'), 138.0 (C-2), 146.9 (C-2'), 149.6 (C-6'), 170.7 (C-13), 180.4 (<u>C</u>OOH), 199.2 (C-11), 206.9 (C-3).

**3,11-Dioxo-2-(pyridine-4-ylmethylideno)-olean-12-en-30-oic** Acid (31). Yield 0.49 g (88%), mp 147–149°C,  $[\alpha]_D^{20}$  +75° (*c* 0.1, CHCl<sub>3</sub>), C<sub>36</sub>H<sub>47</sub>O<sub>4</sub>N (MW 557.76).

PMR spectrum (CDCl<sub>3</sub>, δ, ppm, J/Hz): 0.85, 1.00, 1.10, 1.21, 1.23, 1.35, 1.42 (21H, 7s, 7CH<sub>3</sub>), 1.00–2.70 (19H, m, CH<sub>2</sub>, CH), 5.78 (1H, s, H-12), 7.41 (2H, d, J = 5.3, H-3', H-5'), 8.50 (1H, s, H-7'), 8.65 (2H, d, J = 5.3, H-2', H-6'), 11.0 (1H, br. signal, COO<u>H</u>).

<sup>13</sup>C NMR spectrum (δ, ppm): 15.4, 16.2, 18.1, 18.4, 19.5, 20.6, 22.8, 26.4, 28.5, 31.1, 32.0, 36.5, 37.8, 41.5, 43.3, 43.8, 45.0, 46.0, 48.7, 49.4, 53.8, 54.4, 55.9, 59.2, 124.6 (C-3'), 124.6 (C-5'), 128.3 (C-12), 133.3 (C-2), 138.9 (C-4'), 144.6 (C-7'), 148.8 (C-2'), 148.8 (C-6'), 170.5 (C-13), 179.9 (<u>C</u>OOH), 199.1 (C-11), 206.8 (C-3).

The methods for studying the antituberculosis activity of 7–9, 15–18, 20, 22, 23, 25, 26, 30, and 31 are given at the website www.niaid-aacf.org.

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## REFERENCES

- O. B. Flekhter, O. Yu. Ashavina, E. I. Boreko, L. T. Karachurina, N. I. Pavlova, N. N. Kabal'nova, O. V. Savinova, F. Z. Galin, S. N. Nikolaeva, F. S. Zarudii, L. A. Baltina, and G. A. Tolstikov, *Khim.-farm. Zh.*, **36**, No. 6, 21 (2002).
- L. A. Baltina, O. B. Flekhter, L. R. Nigmatullina, E. I. Boreko, N. I. Pavlova, S. N. Nikolaeva, O. V. Savinova, and G. A. Tolstikov, *Bioorg. Med. Chem. Lett.*, 13, 3549 (2003).
- 3. O. B. Kazakova, G. V. Giniyatullina, E. Y. Yamansarov, and G. A. Tolstikov, *Bioorg. Med. Chem. Lett.*, **20**, 4088 (2010).
- 4. A. Sami, M. Tarua, K. Salme, and Y.-K. Jari, Eur. J. Pharm. Sci., 29, 1 (2006).
- 5. O. B. Flekhter, N. I. Medvedeva, O. S. Kukovinets, L. V. Spirikhin, E. G. Galkin, F. Z. Galin, D. G. Golovanov, N. I. Pavlova, O. V. Savinova, E. I. Boreko, and G. A. Tolstikov, *Bioorg. Khim.*, **6**, 629 (2007).
- 6. O. B. Flekhter, N. I. Medvedeva, G. A. Tolstikov, O. V. Savinova, E. I. Boreko, and F. M. Dolgushin, *Bioorg. Khim.*, **35**, 129 (2009).
- V. E. Kataev, I. Yu. Strobykina, O. V. Andreeva, B. F. Garifullin, R. R. Sharipova, V. F. Mironov, and R. V. Chestnova, *Bioorg. Khim.*, 37, 542 (2011).
- 8. J. Q. Gu, Y. Wang, S. Franzblau, G. Montenegro, D. Yang, and B. N. Timmermann, *Planta. Med.*, 70, 509 (2004).
- 9. S. Suksamrarn, P. Panseeta, S. Kunchanawatta, T. Distaporn, S. Ruktasing, and A. Suksamrarn, *Chem. Pharm. Bull.*, **54**, 535 (2006).
- 10. T. Tanachatchairatana, J. B. Bremner, R. Chokchaisiri, and A. Suksamrarn, Chem. Pharm. Bull., 56, 194 (2008).
- 11. A. Jimenez-Arellanes, M. Meckes, J. Torres, and J. L. Herrera, J. Ethnopharmacol., 111, 202 (2007).
- 12. B. U. Jaki, S. G. Franzblau, L. R. Chadwick, D. C. Lankin, F. Zhang, Y. Wang, and G. F. Pauli, *J. Nat. Prod.*, **71**, 1742 (2008).
- F. Ge, F. Zeng, S. Liu, N. Guo, H. Ye, Y. Song, J. Fan, X. Wu, X. Wang, X. Deng, Q. Jin, and L. Yu, J. Med. Microbiol., 59, 567 (2010).
- 14. A. Jimenez, M. Meckes, V. Alvarez, J. Torres, and R. Parra, *Phytother. Res.*, 19, 320 (2005).
- O. B. Flekhter, E. I. Boreko, L. R. Nigmatullina, E. V. Tret'yakova, N. I. Pavlova, L. A. Baltina, S. N. Nikolaeva,
  O. V. Savinova, V. F. Eremin, F. Z. Galin, and G. A. Tolstikov, *Bioorg. Khim.*, **30**, 89 (2004).
- O. B. Flekhter, E. I. Boreko, L. R. Nigmatullina, N. I. Pavlova, S. N. Nikolaeva, O. V. Savinova, V. F. Eremin, L. A. Baltina, F. Z. Galin, and G. A. Tolstikov, *Bioorg. Khim.*, 29, 326 (2003).
- O. B. Flekhter, L. T. Karachurina, L. R. Nigmatullina, L. A. Baltina, F. S. Zarudii, V. A. Davydova, L. V. Spirikhin, I. P. Baikova, F. Z. Galin, and G. A. Tolstikov, *Khim.-farm. Zh.*, 34, No. 2, 3 (2000).
- O. B. Flekhter, L. R. Nigmatullina, L. A. Baltina, L. T. Karachurina, F. Z. Galin, F. S. Zarudii, G. A. Tolstikov,
  E. I. Boreko, N. I. Pavlova, S. N. Nikolaeva, and O. V. Savinova, *Khim.-farm. Zh.*, 36, No. 9, 26 (2002).
- 19. G. A. Tolstikov, L. A. Baltina, V. P. Grankina, R. M. Kondratenko, and T. G. Tolstikova, *Licorice: Biovariety, Chemistry, and Use in Medicine* [in Russian], Akad. Izd. Geo, Novosibirsk, 2007.
- O. B. Flekhter, E. I. Boreko, L. R. Nigmatullina, N. I. Pavlova, N. I. Medvedeva, S. N. Nikolaeva,
  E. V. Tret'yakova, O. V. Savinova, L. A. Baltina, L. T. Karachurina, F. Z. Galin, F. S. Zarudii, and G. A. Tolstikov, *Khim.-farm. Zh.*, 38, No. 3, 31 (2004).
- 21. N. P. Buu-Hoi, N. D. Xuong, N. H. Nam, F. Binon, and R. Royer, J. Chem. Soc., 1358 (1953).